Immunohistochemistry (IHC) Protocol-Staining Protocol

1. Deparaffinize:

Wash slides with specific reagents in the following order:

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xylene, two times, 5 min each.

100% ethanol, two times, 5 min each
95% ethanol, two times, 5 min each
80% ethanol, once, 5 min
70% ethanol, once, 5 min
50% ethanol, once, 5 min
dH2O, two times, 5 min each
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- 2. (Recommended) Block the endogenous peroxidase activity at room temperature by a 5~10 min incubation in the final developmental 3% H2O2 in distilled water or PBS (pH 7.4).
- 3. Rinse sections with PBS for 5 min.
- 4. Antigen retrieval.
- 5. Rinse sections with PBS for 5 min.
- 6. Apply the blocking antibody (normal goat serum), incubate for 30 min at room temperature, and throw off residual fluid (**don't wash**.).
- 7. Apply the primary antibody 60 min at RT or 4°C for overnight
- 8. Rinse sections 3 times for 5 min each.
- 9. Incubate array slide with a HRP-conjugated secondary antibody at 37°C for 40 min.
- 10. Rinse sections 3 times for 5min each.
- 11. Proceed with chromogen of final developmental DAB or use DAB Kit (Control the degree of staining with regular microscopy).
- 12. Wash sections in distilled water.
- 13. Stain and differentiate slides in hematoxylin.
- 14. Dehydration and transparency of slides.
- 15. Mount slides.